



# STIC Search Report

## Biotech-Chem Library

STIC Database Tracking Number: 10/696023

**TO:** Ralph J Gitomer  
**Location:** 3d65 / 3c18  
**Art Unit:** 1655  
**Friday, March 03, 2006**

**Case Serial Number:** 10/696023

**From:** Noble Jarrell  
**Location:** Biotech-Chem Library  
**Rem 1B71**  
**Phone:** 272-2556

**Noble.jarrell@uspto.gov**

### Search Notes

## SEARCH REQUEST FORM

## Scientific and Technical Information Center

Requester's Full Name: 17 GRITZNER Examiner #: 69630 Date: 2/21/06  
 Art/Unit: 1655 Phone Number 30 \_\_\_\_\_ Serial Number: 10/696,023  
 Mail Box and Bldg/Room Location: \_\_\_\_\_ Results Format Preferred (circle): PAPER DISK E-MAIL  
3C18 3D65

If more than one search is submitted, please prioritize searches in order of need.

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Please provide a detailed statement of the search topic, and describe as specifically as possible the subject matter to be searched. Include the elected species or structures, keywords, synonyms, acronyms, and registry numbers, and combine with the concept or utility of the invention. Define any terms that may have a special meaning. Give examples or relevant citations, authors, etc, if known. Please attach a copy of the cover sheet, pertinent claims, and abstract.

Title of Invention: \_\_\_\_\_

Inventors (please provide full names): \_\_\_\_\_

Earliest Priority Filing Date: \_\_\_\_\_

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Online Time: <u>98</u>		Other <u>      </u>	Other (specify) <u>      </u>

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L69 ANSWER 1 OF 2 HCAPLUS COPYRIGHT 2006 ACS on STN  
AN 2005:1349087 HCAPLUS  
DN 144:83634  
ED Entered STN: 29 Dec 2005  
TI Method for preserving intracellular molecular detail  
IN Davis, Ashley S.; Middleton, Kim M.  
PA USA  
SO U.S. Pat. Appl. Publ., 24 pp.  
CODEN: USXXCO  
DT Patent  
LA English  
IC ICM A01N-0001/02  
INCL 435002000  
CC 9-11 (Biochemical Methods)  
FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
US2005287513	A1	20051229	2003US-0696023	20031030 <--
PRAI 2003US-0696023			20031030	<--

CLASS

PATENT NO.	CLASS	PATENT FAMILY CLASSIFICATION CODES
US 2005287513	ICM	A01N-0001/02
	INCL	435002000
	IPCI	A01N0001-02 [ICM, 7]
	NCL	435/002.000

AB A process and composition is described that allows the operator to fix and preserve tissue culture grown cells such that their intracellular mol. detail is retained for up to four years. This enables increased reproducibility of staining for antigens and small mol. targets for use in the areas of basic research and diagnostic applications.

ST cell preservation biol staining

IT Animal cell line  
(3T3; method for preserving intracellular mol. detail)

IT Animal cell line  
(HT-1080; method for preserving intracellular mol. detail)

IT Animal cell line  
(MCF-7; method for preserving intracellular mol. detail)

IT Fibronectins

RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)  
 (conjugate with rhodamine; method for preserving intracellular mol. detail)

IT Cell junction  
 (focal contact; method for preserving intracellular mol. detail)

IT Immunoassay  
 (immunofluorescent staining; method for preserving intracellular mol. detail)

IT Animal tissue culture  
 Apoptosis  
 Buffers  
 Freeze drying  
 Freezing  
 HeLa cell  
 Mitosis  
 Preservation  
 Preservation solutions (tissue)  
 Staining, biological  
 Storage  
 (method for preserving intracellular mol. detail)

IT Carbohydrates, biological studies  
 Growth factors, animal  
 Lysophosphatidic acids  
 Platelet-derived growth factors  
 RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)  
 (method for preserving intracellular mol. detail)

IT Glass, uses  
 RL: DEV (Device component use); USES (Uses)  
 (method for preserving intracellular mol. detail)

IT Actins  
 RL: PRP (Properties)  
 (method for preserving intracellular mol. detail)

IT Tubulins  
 RL: PRP (Properties)  
 (method for preserving intracellular mol. detail)

IT Culture media  
 (serum deficient; method for preserving intracellular mol. detail)

IT 67-56-1, Methanol, biological studies 111-30-8  
 , Glutaraldehyde 13558-31-1D, derivs., conjugate with  
 fibronectin 219920-04-4, Rhodamine phalloidin  
 RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)  
 (method for preserving intracellular mol. detail)

L69 ANSWER 2 OF 2 HCAPLUS COPYRIGHT 2006 ACS on STN  
 AN 2003:892325 HCAPLUS  
 DN 139:347745  
 ED Entered STN: 14 Nov 2003

TI Preservation of RNA and morphology in cells and tissues  
 IN Vincek, Vladimir; Nassiri, Mehdi; Nadji, Mehrdad; Morales, Azorides R.  
 PA USA

SO U.S. Pat. Appl. Publ., 12 pp.  
 CODEN: USXXCO

DT Patent

LA English

IC ICM A01N-0001/02  
 ICS C12N-0005/00

INCL 435001100; 435374000; 436008000

CC 9-11 (Biochemical Methods)

Section cross-reference(s): 14

FAN.CNT 1

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
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PI US2003211452	A1	20031113	2002US-0141780	20020510

PRAI 2002US-0141780

20020510

## CLASS

## PATENT NO. CLASS PATENT FAMILY CLASSIFICATION CODES

US 2003211452	ICM	A01N-0001/02
	ICS	C12N-0005/00
	INCL	435001100; 435374000; 436008000
	IPCI	A01N0001-02 [ICM, 7]; C12N0005-00 [ICS, 7]
	IPCR	A01N0001-00 [I,A]; A01N0001-00 [I,C]
	NCL	435/001.100
	ECLA	A01N001/00

AB A solution for preservation and/or storage of a cell or tissue is described. This simple nonaq. composition can have 10% polyethylene glycol and 90% methanol. It can be used at room temperature. Special chems., equipment, and techniques are not needed. Tissue preserved with and/or stored in the solution can be processed for cytol. or histol., including chemical staining and/or antibody binding, by a variety of methods; antigen, DNA, and RNA can be extracted from processed tissue in high yield and with minimal or no degradation. Advantages of the solution include: economy and safety, easy access to archival material, and compatibility with both cellular and genetic analyses. The use and manufacture of the solution are also described.

ST preservation RNA morphol cell tissue

IT Uterus, neoplasm

(leiomyoma; preservation of RNA and morphol. in cells and tissues)

IT Animal tissue culture

Cell

Human

Kidney, disease

Neoplasm

Preservation

Staining, biological

(preservation of RNA and morphol. in cells and tissues)

IT RNA

RL: BSU (Biological study, unclassified); BIOL (Biological study)

(preservation of RNA and morphol. in cells and tissues)

IT Polyoxyalkylenes, biological studies

RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)

(preservation of RNA and morphol. in cells and tissues)

IT Antigens

DNA

RL: PEP (Physical, engineering or chemical process); PYP (Physical process); PROC (Process)

(preservation of RNA and morphol. in cells and tissues)

IT Infection

Inflammation

Kidney, disease

(pyelonephritis; preservation of RNA and morphol. in cells and tissues)

IT 9001-99-4

RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)

(A; preservation of RNA and morphol. in cells and tissues)

IT 64-17-5, Ethanol, biological studies 67-56-1, Methanol, biological studies 67-63-0, Isopropanol, biological studies 67-64-1, Acetone, biological studies 67-66-3, Chloroform, biological studies 111-30-8, Glutaraldehyde 1330-20-7, Xylene, biological studies 7722-84-1, Hydrogen peroxide, biological studies 9003-99-0, Peroxidase 25322-68-3, Polyethylene glycol 650607-24-2, Methacarn

RL: BUU (Biological use, unclassified); BIOL (Biological study); USES (Uses)

(preservation of RNA and morphol. in cells and tissues)

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FILE 'WPIX' ENTERED AT 16:31:16 ON 03 MAR 2006  
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FILE LAST UPDATED: 2 MAR 2006 <20060302/UP>  
 MOST RECENT DERWENT UPDATE: 200615 <200615/DW>  
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[<<<](http://scientific.thomson.com/media/scpdf/ ipcrdwpi.pdf)  
 'BIX' IS DEFAULT SEARCH FIELD FOR 'WPIX' FILE

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L124 ANSWER 1 OF 9 WPIX COPYRIGHT 2006 THE THOMSON CORP on STN  
 AN 2006-055998 [06] WPIX  
 DNC C2006-020928

TI Preserving tissue culture cells on glass slides by fixing with  
 glutaraldehyde or methanol, followed by using  
 preservative agent, followed by rapid freeze step, and lyophilization and  
 storage under cool and desiccated conditions.

DC B04 D16  
 IN DAVIS, A S; MIDDLETON, K M  
 PA (DAVI-I) DAVIS A S; (MIDD-I) MIDDLETON K M  
 CYC 1  
 PI US--2005287513 A1 20051229 (200606)\* 24 A01N-001-02 <--  
 ADT US--2005287513 A1 2003US-0696023 20031030  
 PRAI 2003US-0696023 20031030

IC ICM A01N-001-02  
 AB US2005287513 A UPAB: 20060124  
 NOVELTY - Preserving (M1) tissue culture cells on glass slides, comprising  
 fixing with glutaraldehyde or methanol, using a  
 preservative agent containing a buffer, a sugar and a carbohydrate  
 polymer, using a rapid freeze step, lyophilizing and storing under cool  
 and desiccated conditions, is new.

DETAILED DESCRIPTION - An INDEPENDENT CLAIM is also included for  
 using rhodamine fibronectin as a rapid stain for focal adhesion plaques.

USE - (M1) is useful for preserving tissue culture cells on glass  
 slides. (M1) is suitable for Swiss 3T3 cells, HT1080 cells, HeLa cells,  
 MCF-7 cells, other cell lines, mitotic cell preparations, apoptotic cell  
 preparations, growth factor treated cells, lysophosphatidic acid treated  
 cells, platelet derived growth factor treated cells, tumor necrosis factor  
 alpha treated cells, serum starved cells, or probing of focal adhesion  
 plaques (claimed). (M1) is useful in basic research and diagnostic  
 applications, and in cell biology for detecting mitotic cell.

ADVANTAGE - (M1) preserves long lasting cell samples for immunological examination. (M1) allows the operator to fix and preserve tissue culture grown cells so that their intracellular molecular detail is retained for up to four years. (M1) enables increased reproducibility of staining for antigens and small molecule targets.

DESCRIPTION OF DRAWING(S) - The figure shows schematic diagram of the process of making preserved cells.

Dwg.1/14

FS CPI  
FA AB; GI; DCN  
MC CPI: B04-F01; B04-H19; B06-A02; B06-A03; B10-D01; B10-E04D;  
B11-C07B1; B11-C08E1; B12-K04; D05-H08; D05-H09

ABEX UPTX: 20060124

EXAMPLE - No relevant example is given.

TECH UPTX: 20060124

TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: (M1) results in retention of nanometer scale molecular structure detail. (M1) results in a product that has a shelf life greater than four years at 4 degreesC. (M1) produces a preparation of cells on a glass slide.

L124 ANSWER 2 OF 9 WPIX COPYRIGHT 2006 THE THOMSON CORP.on STN

AN 2005-628188. [64] WPIX

DNC C2005-188430

TI Composition; useful for preserving biological specimens suspected to contain circulating tumor cells, consists of an anti-coagulating agent and a stabilizing agent.

DC A25 A89 B04 D22 J04

IN HERMAN, M; RAO, G C; RUTNER, H; TERSTAPPEN, L W M M

PA (HERM-I) HERMAN M; (RAOG-I) RAO G C; (RUTN-I) RUTNER H; (TERS-I) TERSTAPPEN L W M M

CYC 1

PI US--2005181353 A1 20050818 (200564)\* 19 A01N-001-02 <--

ADT US--2005181353 A1 2004US-0780349 20040217

PRAI 2004US-0780349 20040217

IC ICM A01N-001-02

ICS A61K-031-198; A61K-031-727

AB US2005181353 A UPAB: 20051006

NOVELTY - Composition (A) for preserving biological specimens comprising an anti-coagulating agent and a stabilizing agent, is new.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for:

(1) a stabilized cell composition (B) consisting of a biological specimen, an anti-coagulating agent, and a stabilizing agent;

(2) preserving biological specimens consisting of obtaining a biological specimen that contains cells, and contacting the biological specimen with a stabilizing agent capable of stabilizing the cells; and

(3) an apparatus for preserving biological specimens consisting of an evacuated blood drawn tube, and anti-coagulating agent and a stabilizing agent.

USE - (A) is useful for preserving blood samples suspected to contain circulating tumor cells (claimed), prior to analysis. The stabilizers are useful to inhibit damage to magnetically labeled cells that occur both for magnetic and non-magnetic stresses even during normal specimen processing (centrifugation, vortexing and pipetting).

ADVANTAGE - (A) is more effective in stabilizing the circulating tumor cells.

Dwg.0/6

FS CPI

FA AB; DCN

MC CPI: A12-L04; B04-B04D; B04-C02E1; B04-C03C;  
B04-F02A; B05-A03A1; B05-A03A4; B05-A03B; B10-A13D; B10-B01B;  
B10-C02; B10-D01; B11-C06; D09-A03; J04-C01

TECH UPTX: 20051006

TECHNOLOGY FOCUS - BIOLOGY - Preferred Components: The anti-coagulating agent is a chelating agent (ethylenediamine tetraacetic acid (EDTA), diethylenetriamine pentaacetic acid (DTPA), 1,2-diaminocyclohexane tetraacetic acid (DCTA) or ethylenbis(oxyethylenenitrilo) tetraacetic

acid (EGTA)) or a complexing agent (heparin and citrate). The stabilizing agent is a formaldehyde donor (methyol or hydroxymethyl derivatives of amines or amides, diazolinidinyl urea, imidazolidinyl urea, methenamine or paraformaldehyde) or an aldehyde (formaldehyde, glutaraldehyde or glyoxal) combined with at least one heavy metal element (chromium, manganese or zinc). (A) comprises an additional stabilizing agent (polyethylene glycol). The molecular weight of the polyethylene glycol is 1000-35000 (8000-20000).

Preferred Method: The biological specimen is a fraction of blood suspected to contain circulating tumor cells. The circulating tumor cells have been stabilized by the stabilizing agent. In (B), the anti-coagulating and stabilizing agents are about 0.1-50% (preferably 0.3-5%) of the total volume of the biological specimen. In the method of preserving biological sample, the specimen is further contacted with the anti-coagulating agent. The anti-coagulating agent and the stabilizing agent are combined before contacting the biological specimen.

L124 ANSWER 3 OF 9 WPIX COPYRIGHT 2006 THE THOMSON CORP on STN  
 AN 2005-403950 [41] WPIX  
 CR 2003-341615 [32]; 2003-864354 [80]; 2004-804426 [79]; 2005-030768 [03]  
 DNN N2005-327674 DNC C2005-124819  
 TI Preparation of a fixed biological tissue, useful for implantation/transplantation, comprises harvesting, shaping or sterilizing the biological tissue.  
 DC B07 D22 P32  
 IN SARAC, T P  
 PA (CLEV-N) CLEVELAND CLINIC FOUND  
 CYC 1  
 PI US--2005119728 A1 20050602 (200541)\* 20 A61F-002-06  
 ADT US--2005119728 A1 Div ex 2001US-0908764 20010719, CIP of 2003US-0409884  
 20030409, 2005US-0029687 20050105  
 FDT US--2005119728 A1 Div ex US-----6579307  
 PRAI 2005US-0029687 20050105; 2001US-0908764 20010719;  
 2003US-0409884 20030409  
 IC ICM A61F-002-06  
 AB US2005119728 A UPAB: 20050629  
 NOVELTY - Preparing a fixed biological tissue (A) comprising harvesting at least one layer of (A); shaping (A); at least partially cross-linking the shaped (A); sterilizing cross-linked (A) and inactivating prions in the sterilized (A), is new.  
 USE - The method is useful for tissue fixation and to methods of treating tissues useful in implantation/transplantation.  
 Dwg.0/15  
 FS CPI GMPI  
 FA AB; DCN  
 MC CPI: B04-C02B; B04-C03C; B04-F01; B04-N10; B10-D01;  
 B10-E04D; B11-C04A; D09-A03; D09-C01  
 TECH UPTX: 20050629  
 TECHNOLOGY FOCUS - PHARMACEUTICALS - Preferred Method: Shaping of (A) comprises cutting the harvested layer of (A) to a desired size and suturing the cut (A) to a member. (A) is pericardial tissue, peritoneal tissue or pleural tissue. (A) is at least partially cross-linked by contacting the biological tissue with a solution containing at least one cross-linking agent. The cross-linking agent is glutaraldehyde, formaldehyde, glyceraldehyde, paraformaldehyde and/or dialdehyde. (A) is sterilized by contacting (A) with a solution containing an alcohol (ethanol, methanol, propanol and/or isopropanol) at a concentration of 10-90 vol.%. The prions in (A) are inactivated by contacting (A) with a basic solution. The basic solution having a molar concentration of 0.5-4.0 M. The method of preparing (A) further comprises contacting (A) with at least one solution that causes lysis of cells of (A) and applying at least one pharmacological agent to (A). The solution that causes cell lysis is a hypertonic solution that has a salt concentration of 2-8 wt.%. The method of preparing (A) further comprises applying at least one pharmacological agent (comprises at least one of an anti-inflammatory agent, an anti-proliferative agent, an anti-coagulation

agent or an anti-platelet aggregation agent) to the fixed biological tissue. The method of preparing (A) further comprises contacting the shaped (A) tissue with a cell lysis solution prior to cross-linking (A). The biological tissue is at least partially cross-linked by contacting (A) with a solution containing at least one cross-linking agent such as glutaraldehyde, formaldehyde, glyceraldehyde, paraformaldehyde and/or dialdehyde starch. The preparation of (A) further comprises applying least one pharmacological agent to the fixed tissue. The layer of (A) comprises a plurality of cells.

Li24 ANSWER 4 OF 9 WPIX COPYRIGHT 2006 THE THOMSON CORP on STN

AN 2004-430865 [40] WPIX

DNC C2004-161238

TI Preserving eukaryotic cell, tissue or organ, involves contacting cell, tissue or organ with hypothermic storage solution, and vitrifying cell, tissue or organ within cell, tissue or organ and in hypothermic storage solution.

DC A96 B04 C06 D16

IN BAUST, J G; BAUST, J M; MATHEW, A J; VANBUSKIRK, R  
PA (BIOL-N) BIOLIFE SOLUTIONS INC

CYC 105

PI US--2004096813 A1 20040520 (200440)\* 10 A01N-001-02 <--  
WO--2004046308 A2 20040603 (200440) EN C12N-000-00

RW: AT BE BG CH CY CZ DE DK EA EE ES FI FR GB GH GM GR HU IE IT KE LS  
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W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK  
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KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NI NO NZ OM PG PH  
PL PT RO RU SC SD SE SG SK SL SY TJ TM TN TR TT TZ UA UG UZ VC VN  
YU ZA ZM ZW

AU--2003273276 A1 20040615 (200470) A01N-001-02 <--  
US-----6921633 B2 20050726 (200549) A01N-001-00 <--

ADT US--2004096813 A1 2002US-0298497 20021118; WO--2004046308 A2  
2003WO-US027688 20030902; AU--2003273276 A1 2003AU-0273276 20030902;  
US-----6921633 B2 2002US-0298497 20021118

FDT AU--2003273276 A1 Based on WO--2004046308

PRAI 2002US-0298497 20021118

IC ICM A01N-001-00; A01N-001-02; C12N-000-00

AB US2004096813 A UPAB: 20040624

NOVELTY - Preserving a eukaryotic cell, tissue or organ, involves contacting the cell, tissue or organ with hypothermic storage solution comprising a composition that inhibits apoptosis, and a concentration of a vitrification composition, where the vitrification occurs both within the cell, tissue or organ and in the hypothermic storage solution comprising and comprised by the cell, tissue or organ.

DETAILED DESCRIPTION - Preserving (M1) a eukaryotic cell, tissue or organ, involves contacting the cell, tissue or organ with hypothermic storage solution, where the solution comprises a composition that inhibits apoptosis, and a concentration of a vitrification composition that is sufficient for vitrification of the solution, and vitrifying the cell, tissue or organ, where the vitrification occurs both within the cell, tissue or organ and in the hypothermic storage solution comprising and comprised by the cell, tissue or organ.

An INDEPENDENT CLAIM is also included for a hypothermic preservation solution comprising, a composition that inhibits apoptosis, and a vitrification composition that comprises a concentration of one or more agents that is sufficient for vitrification of the solution when the temperature of the solution is reduced below the glass transition temperature (Tg) of the solution.

USE - (M1) is useful for preserving a eukaryotic cell, tissue or organ (claimed). The organs which can be preserved prior to transplantation in a recipient patient by (M1) include lung, liver, heart, kidney, gut, eye and skin. Tissue such as bone marrow, and cells such as erythrocytes and leukocytes are also preserved by (M1).

ADVANTAGE - (M1) enables to preserve cells, tissues or organs for long periods without significant loss of viability. (M1) enables long-term

preservation of gametes and embryos for reproductive procedures such as in vitro fertilization.

Dwg.0/0

FS CPI

FA AB; DCN

MC CPI: A12-V03; B03-A; B03-F; B03-G; B03-H; B04-C02C; B04-C03; B04-D01; B04-F01; B04-L03; B05-C03; B07-A02A; B07-A02B; B10-A07; B10-A11B; B10-A17; B10-B02D; B10-C04E; B10-D01; B10-D03; B10-E04C; B10-E04D; B14-D07; B14-D10; C03-A; C03-F; C03-G; C03-H; C04-C02C; C04-C03; C04-D01; C04-F01; C04-L03; C05-C03; C07-A02A; C07-A02B; C10-A07; C10-A11B; C10-A17; C10-B02D; C10-C04E; C10-D01; C10-D03; C10-E04C; C10-E04D; C14-D07; C14-D10; D05-A02A

ABEX UPTX: 20040624

EXAMPLE - No relevant example given.

TECH UPTX: 20040624

TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: In (M1), the hypothermic storage solution has a glass transition temperature lower than its homogenous nucleation temperature. The vitrification composition comprises one or more agents chosen from sucrose, trehalose, lactose, glucose, dimethyl sulfoxide (DMSO), propylene glycol, ethylene glycol, dextran, glycerol, hydroxyethyl starch, polyvinyl pyrrolidone, formamide, 1,2-propanediol, ethanol, methanol and polyethylene oxide. The composition that inhibits apoptosis comprises an agent that interacts with a polypeptide that participates in an apoptotic pathway. The agent inhibits the activity of the polypeptide, maintains or potentiates the activity of the polypeptide.

The agent is chosen from caspase inhibitor, calpain inhibitor and an inhibitor of nitrous oxide synthase. The agent is a caspase inhibitor and is chosen from peptide fluoromethyl ketone, CHO, peptide chloromethyl ketone, 2,6-dichlorobenzoyloxymethyl ketone (DCB), 2,6-dimethylbenzoyloxymethyl ketone (AOM) and 2,6-bis(trifluoromethyl)benzoyloxymethyl ketone (FAOM). The agent is a calpain inhibitor and is chosen from Leupeptin, Calpain inhibitors I, II, III, IV and V, calpeptin, Loxastatin, a peptide chloromethyl ketone and a peptide fluoromethyl ketone.

The composition that inhibits apoptosis comprises an antioxidant. The antioxidant is chosen from glutathione, N-acetyl cysteine, beta carotene, vitamins E, D, C and A, nitric oxide, L-arginine and super oxide dismutase. The composition that inhibits apoptosis comprises an agent chosen from free radical scavenger, zinc chelator and a calcium chelator. The agent is preferably a free radical scavenger. The free radical scavenger is chosen from vitamins E, D, C and A, nitric oxide, L-arginine and super oxide dismutase.

L124 ANSWER 5 OF 9 WPIX COPYRIGHT 2006 THE THOMSON CORP on STN

AN 2004-154374 [15] WPIX

DNC C2004-061369

TI Cell or tissue storage composition, for e.g. used in isolating nucleic acid, comprises non-aqueous solution comprising polyethylene glycol, and methanol.

DC A96 B04 D16

IN MORALES, A R; NADJI, M; NASSIRI, M; VINCEK, V

PA (MORA-I) MORALES A R; (NADJ-I) NADJI M; (NASS-I) NASSIRI M; (VINC-I) VINCEK V

CYC 1

PI US--2003211452 A1 20031113 (200415)\* 12 A01N-001-02 <--

ADT US--2003211452 A1 2002US-0141780 20020510

PRAI 2002US-0141780 20020510

IC ICM A01N-001-02

ICS C12N-005-00

AB US2003211452 A UPAB: 20040302

NOVELTY - A cell or tissue storage composition comprises non-aqueous solution comprising 5-20% polyethylene glycol (PEG) and 80-95% methanol.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for:

(a) making the composition comprising mixing PEG and methanol

; and

(b) a cell or tissue holder containing the composition and adapted to hold cell and/or tissue.

USE - The composition is used for preserving cell or tissue by contacting the composition with the cell or tissue within 30 minutes, and/or storing cell or tissue by contacting the composition with the cell or tissue for at least 2 weeks. It is used in isolating nucleic acid i.e. RNA. (All claimed).

ADVANTAGE - The inventive composition may be used in room temperature without need special chemicals, equipment or techniques. It is capable of extracting DNA and RNA in high yield with minimal or no degradation. It is economic, safe, easy to access to archival material, and compatible with both cellular and genetic analyses.

Dwg. 0/2

FS CPI

FA AB; DCN

MC CPI: A05-H03A3; A12-V02; A12-V03; B04-C03C; B04-E01; B04-F01;  
B10-E04D; B11-B; D05-H01; D05-H02; D05-H08; D05-H12; D05-H13

ABEX UPTX: 20040302

EXAMPLE - 50 mg Fresh tissue persevered in 10% polyethylene glycol (PEG) and 90% methanol mixture, was plated in 1 ml Trizol reagent. The samples were incubated at room temperature for 5 minutes and 0.2 ml chloroform was added by hand mixing for 15 seconds. Samples were centrifuged at 12000 g for 15 minutes at 5 degrees C. Aqueous phase was removed and precipitated using 0.5 ml of isopropyl alcohol. After 10 minutes of incubation at room temperature, samples were cooled to 5 degrees C and centrifuged at 12000 g for 10 minutes. The RNA pellet was washed in 70% ethanol, air dried for 15 minutes, and dissolved in 100 microl ribonuclease-free water.

TECH UPTX: 20040302

TECHNOLOGY FOCUS - POLYMERS - Preferred Composition: The non-aqueous solution comprises 10-15 (preferably 10)% polyethylene glycol (PEG) and 85-90 (preferably 90)% methanol.

Preferred Parameters: The PEG has a molecular weight at most 600 (preferably at most 400) Daltons.

TECHNOLOGY FOCUS - BIOTECHNOLOGY - Preferred Method: The method further comprises extracting nucleic acid from at least a portion of the cell or tissue.

L124 ANSWER 6 OF 9 WPIX COPYRIGHT 2006 THE THOMSON CORP on STN

AN 2003-354443 [33] WPIX

CR 2003-278691 [27]

DNN N2003-283266 DNC C2003-093346

TI Composition for preserving biological specimens or blood samples suspected to contain circulating tumor cells, comprises anti-coagulating agent and stabilizing agent.

DC A96 B04 D16 E19 S03

IN HERMANN, M; RAO, G C; RUTNER, H; TERSTAPPEN, L; FOULK, B; OHARA, S M;  
ZWEITZIG, D

PA (IMMU-N) IMMUNIVEST CORP; (FOUL-I) FOULK B; (OHAR-I) OHARA S M; (ZWEI-I)  
ZWEITZIG D

CYC 101

PI WO--2003018757 A2 20030306 (200333)\* EN 38 C12N-000-00

RW: AT BE BG CH CY CZ DE DK EA EE ES FI FR GB GH GM GR IE IT KE LS LU  
MC MW MZ NL OA PT SD SE SK SL SZ TR TZ UG ZM ZW

W: AE AG AL AM AT AU AZ BA BB BG BR BY BZ CA CH CN CO CR CU CZ DE DK  
DM DZ EC EE ES FI GB GD GE GH GM HR HU ID IL IN IS JP KE KG KP KR  
KZ LC LK LR LS LT LU LV MA MD MG MK MN MW MX MZ NO NZ OM PH PL PT  
RO RU SD SE SG SI SK SL TJ TM TN TR TT TZ UA UG US UZ VN YU ZA ZM  
ZW

EP----1425383 A2 20040609 (200438) EN C12N-001-00

R: AL AT BE BG CH CY CZ DE DK EE ES FI FR GB GR IE IT LI LT LU LV MC  
MK NL PT RO SE SI SK TR

BR---200212124 A 20040720 (200451) A01N-001-02 <--

AU--2002331696 A1 20030310 (200452) C12N-000-00

JP--2005501236 W 20050113 (200506) 68 G01N-033-48  
 US--2006008807 A1 20060112 (200605) C12Q-001-68  
 ADT WO--2003018757 A2 2002WO-US026867 20020823; EP----1425383 A2  
 2002EP-0768676 20020823, 2002WO-US26867 20020823; BR---200212124 A  
 2002BR-0012124 20020823, 2002WO-US26867 20020823; AU--2002331696 A1  
 2002AU-0331696 20020823; JP--2005501236 W 2002WO-US26867 20020823,  
 2003JP-0523608 20020823; US--2006008807 A1 2004US-0826585 20040416  
 FDT EP----1425383 A2 Based on WO--2003018757; BR---200212124 A Based on  
 WO--2003018757; AU--2002331696 A1 Based on WO--2003018757; JP--2005501236  
 W Based on WO--2003018757  
 PRAI 2002US-369628P 20020403; 2001US-314151P 20010823  
 IC ICM A01N-001-02; C12N-000-00; C12N-001-00; C12Q-001-68;  
 G01N-033-48  
 AB WO2003018757 A UPAB: 20060120

NOVELTY - A composition (I) for preserving biological specimens or blood samples suspected to contain circulating tumor cells, consists of an anti-coagulating agent, and a stabilizing agent.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for the following:

(1) a stabilized cell composition (II) consisting of a biological specimen, an anti-coagulant agent, and a stabilizing agent; and

(2) an apparatus for preserving blood specimen or blood samples suspected to contain circulating tumor cells, consisting of an evacuated blood draw tube containing an anti-coagulating agent and a stabilizing agent.

USE - (I) is useful for preserving biological specimens or blood samples suspected to contain circulating tumor cells (claimed). (I) is useful for accurate analysis of circulating tumor cells (CTC) which serve as diagnostically important indicators of tumor burden, the proliferative potential of tumor cells and/or the effectiveness of therapy.

ADVANTAGE - (I) minimizes losses of target cells and formation of debris and aggregates from target cells, non-target cells and plasma components, thus allowing more accurate analysis and classification of CTC and ultimately of tumor burdens in cancer patients. The stabilizers inhibit damage to magnetically labeled cells that may occur both from magnetic and non-magnetic stresses even during normal specimen processing such as centrifugation, vortexing and pipetting.

Dwg.0/6

FS CPI EPI

FA AB; DCN

MC CPI: A12-L04; A12-V03C2; B04-B04D5; B04-C02E1;  
 B04-C03C; B05-A03; B06-D17; B07-D09; B10-B02; B10-C02; B10-D01;  
 B11-C08E; B12-K04A1; D05-H09; E05-L03A; E05-L03C; E06-D17; E07-D09D;  
 E10-B01A2; E10-B01C1; E10-B03A2; E10-B03B2; E10-C04A; E10-D01D;  
 E10-D03

EPI: S03-E13A; S03-E14H6

ABEX UPTX: 20030526

EXAMPLE - The effect of different stabilizers on recovery of circulating tumor cells (CTC) after enrichment from 24-hour-old specimens was examined. Blood samples from patients with advanced carcinomas were obtained and treated with stabilizers within 2 hours of blood draw. Blood drawn from each patient into different tubes containing EDTA was pooled, and equal volumes were aliquoted into separate tubes. Various additives, consisting of Cyto-Chex, StabilCyte, and TRANSfix stabilizer, were added to the separate tubes at 30% Cyto-Chex stabilizer, 20% StabilCyte stabilizer, and 10% TRANSfix stabilizer. One tube was used as the control to which no buffer or stabilizer was added. The samples were then mixed and stored at room temperature for 24 hours. An equal volume of Immunicon System Buffer (PBS containing 0.5% BSA, 0.2% casein and 0.1% sodium azide) was added to each sample. After mixing, the samples were centrifuged at 800x for 10 minutes to remove plasma. Immunicon AB buffer (system buffer containing streptavidin as a mediator of controlled reversible aggregation) was added to each tube to final volume of 1.5x the initial blood volume. After mixing the samples, CA EpCAM ferrofluid (0.2  $\mu$ m magnetic particles coupled to anti-EpCAM antibody and with desthiobiotin as a mediator of controlled reversible aggregation) was added to the

samples to magnetically label CTC. The samples were incubated with the CA EpCAM ferrofluid inside Immunicon quadruple high-gradient magnetic separators, for two 10 minute time periods with remixing outside the QMS17 magnets after each 10 minutes time period. After these magnetic incubations, the samples were magnetically separated for 20 minutes. The uncollected fractions were aspirated, and the tubes were removed from the QMS17. The magnetically collected fractions were resuspended in System buffer and re-separated in the QMS17 for 10 minutes. The uncollected fractions were aspirated again and the collected cells were resuspended in 200 mul of Immuniperm to permeabilize the captured cells to allow intracellular staining. The permeabilized samples were stained for 15 minutes with a cocktail consisting of several fluorescent markers: 20 mul anti-cytokeratin-PE, 20 mul anti-CD45-APC and 20 mul DAPI. Anti-cytokeratin stains epithelial cells and anti-CD45 stains leukocytes to differentiate any non-specifically stained leukocytes from target CTC. DAPI was used to identify all nucleated cells and to differentiate cells from non-nucleated cell debris. After washing out excess staining reagents by magnetic separation, the samples were resuspended in 320 mul Immunicon CellFix. Each sample was transferred to an Immunicon CellSpotter chamber. Higher numbers of intact and suspect CTC were detected in most of the patient samples after 24-hour storage of these blood samples with all three stabilizers when compared to blood samples with no added stabilizer. These data clearly showed that addition of stabilizers to blood samples from cancer patients preserve CTC during storage and in the sample processing steps, thus permitting more accurate classification of in vivo levels of intact CTC and suspect CTC.

TECH

UPTX: 20030526

TECHNOLOGY FOCUS - ORGANIC CHEMISTRY - Preferred Composition: The anti-coagulating agent is a chelating agent such as ethylenediamine tetraacetic acid (EDTA), diethylenetriamine pentaacetic acid (DTPA), 1,2-diaminocyclohexane tetraacetic acid (DCTA) or ethylenebis(oxyethylenenitrilo) tetraacetic acid (EGTA), or a complexing agent such as heparin or citrate. The stabilizing agent is a formaldehyde donor such as methylol or hydroxymethyl derivatives of amines or amides, diazolinidinyl urea, imidazolidinyl urea, methenamine, or paraformaldehyde; an aldehyde such as formaldehyde, glutaraldehyde or glyoxal; their combination with at least one heavy metal element such as chromium, manganese or zinc. The additional stabilizing agent is polyethylene glycol of molecular weight 1000-35000, preferably 5000-20000 or 8000-20000. In (II), the biological specimen is a fraction of blood suspected to contain circulating tumor cells which have been stabilized by the stabilizing agent. The anti-coagulating agents and the stabilizing agents are present in volumes of 0.1-5%, 0.3-30% or preferably 0.3-5% of the total volume of the biological specimen. The additional stabilizing agent is polyethylene glycol at a concentration of 0.1-5%, preferably 0.1-0.5% of the specimen volume.

L124 ANSWER 7 OF 9 WPIX COPYRIGHT 2006 THE THOMSON CORP on STN

AN 1997-261095 [24] WPIX

CR 1992-367318 [45]

DNC C1997-084517

TI Cell sample preservation solution for cytological samples - contains water-miscible alcohol, magnesium and calcium ions and buffer, preserves cell nuclear morphology.

DC B04 D22

IN HURLEY, A A; LAPEN, D C; OUD, P S

PA (CYTY-N) CYTYC CORP

CYC 10

PI EP-----772972 A1 19970514 (199724)\* EN 9 A01N-001-02 <--  
R: BE CH DE DK FR GB IT LI NL SEEP-----772972 B1 20021002 (200272) EN A01N-001-02 <--  
R: BE CH DE DK FR GB IT LI NL SE

DE-----69133125 E 20021107 (200281) A01N-001-02 &lt;--

ADT EP-----772972 A1 Div ex 1991EP-0110960 19910702, 1996EP-0120872 19910702;

EP-----772972 B1 Div ex 1991EP-0110960 19910702, 1996EP-0120872 19910702;

DE-----69133125 E 1991DE-0633125 19910702, 1996EP-0120872 19910702

FDT EP-----772972 B1 Div ex EP-----511430; DE----69133125 E Based on

EP-----772972

PRAI 1991US-0694452 19910501

REP 1.Jnl.Ref; BE---906129; EP---237863; FR---2490928

IC ICM A01N-001-02

AB EP 772972 A UPAB: 20021216

An aqueous alcohol-buffer solution (I), for preservation of the nuclear morphology of cells, comprises: (A) a water-miscible alcohol; (B) magnesium and calcium ions in an amount sufficient to preserve nuclear morphology, and (C) a buffering agent which maintains the solution with the cells at pH 2-7.

(A) is methanol and forms ca. 20% of (I). (C) is an acetate buffer. (I) further contains a sodium salt (preferably sodium chloride, especially at 0.1% of (I)) and a potassium salt (especially potassium chloride) in amounts sufficient to stabilise the cells. (I) especially contains methanol, magnesium acetate, calcium acetate, sodium chloride and potassium chloride.

USE - (I) is a cell fixing solution useful for preserving cell or tissue samples (e.g vaginal or muscle cells from a patient) at ambient temperature, prior to cytological or histological analysis. The samples may be preserved for relatively long periods or for transport.

ADVANTAGE - (I) maintains cell membranes intact for subsequent staining, destroys microbial pathogens (e.g. *Candida albicans*, *Aspergillus niger*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Staphylococcus aureus*), inhibits retroviral activity and (in some cases) removes undesirable protein material from the solution. Typically cells can be preserved for at least 3 weeks at 4-38 deg. C, while retaining sufficient structure to allow staining without significant loss of integrity.

Dwg.0/0

FS CPI

FA AB

MC CPI: B04-F01; D09-A01C

L124 ANSWER 8 OF 9 WPIX COPYRIGHT 2006 THE THOMSON CORP on STN

AN 1993-003401 [01] WPIX

DNC C1993-001507

TI Blood smear preparation - by immediate fixation with glutaraldehyde after sampling.

DC B04 C07

PA (YAWA) NIPPON STEEL CORP; (TAIF) TAIYO FISHERY CO LTD

CYC 1

PI JP----04330001 A 19921118 (199301)\* 4 A01N-001-02 <--

ADT JP----04330001 A 1991JP-0100211 19910501

PRAI 1991JP-0100211 19910501

IC ICM A01N-001-02

AB JP 04330001 A UPAB: 19931118

Blood smear preparation is carried out by fixing with a fixing agent, pref. glutaraldehyde, immediately after sampling.

USE/ADVANTAGE - The method permits easy preparation, without relying on a skilled technician. Blood smears of stable quality with a definite contour of blood cells are obtd.

In an example, blood was sampled using a syringe from the caudal part of cultured yellowtail (2-year-old fish weighing 2kg). Immediately after sampling, the blood was dropped on a 0.5% glutaraldehyde solution in order to fix the blood and prepare the smear. This was then Giemsa-stained, and blood cells observed with a microscope. In the meantime, another specimen was fixed by the ordinary procedure and then stained the same way. The 0.5% glutaraldehyde-fixed preparation showed clearly demarcated blood cells while the ordinary preparation had a poor blood cell contour

Dwg.0/0

FS CPI

FA AB; DCN

MC CPI: B04-B04D1; C04-B04D1; B04-B04D5;  
C04-B04D5; B10-D01; C10-D01; B11-C07B1; C11-C07B1; B12-K04A;  
C12-K04A

L124 ANSWER 9 OF 9 WPIX COPYRIGHT 2006 THE THOMSON CORP on STN  
 AN 1977-27173Y [16] WPIX  
 TI Tanned blood platelets and preparation - by treatment with (5)-sulpho-  
 salicylic acid or glutaraldehyde, for use in diagnostic tests  
 for antibodies.

DC A96 B04 S03 S05  
 PA (BEHW) BEHRINGWERKE AG

CYC 13

PI BE-----847354 A 19770415 (197716)\*  
 DE-----2546166 A 19770428 (197718)  
 NL-----7611152 A 19770419 (197718)  
 JP-----52047916 A 19770416 (197721)  
 SE-----7611505 A 19770509 (197721)  
 DK-----7604637 A 19770613 (197727)  
 FR-----2328189 A 19770617 (197729)  
 US-----4157383 A 19790605 (197925)  
 AT-----7607656 A 19790715 (197931)  
 GB-----1554886 A 19791031 (197944)  
 CA-----1077394 A 19800513 (198022)  
 CH-----622672 A 19810430 (198120)  
 IT-----1069039 B 19850321 (198531)

PRAI 1975DE-2546166 19751015

IC A01N-001-00; A61K-035-14; C07C-000-00; G01N-001-28; G01N-031-00;  
 G01N-033-16

AB BE 847354 A UPAB: 19930901

Tanning is preferably with 5-sulphosalicylic acid or glutaric aldehyde at a concentration of 0.002-5% w/v. It is pref. to protect the platelets with a lyophilic colloid, e.g. polyvinylpyrrolidone. The novel tanned platelets are stable and do not tend to aggregate. When suspended in a protective colloid solution they may be kept at 4 degrees C for several months.

Freeze-dried tanned platelets may be kept much longer.

Typically, freshly centrifuged blood platelets are mixed with pH 7.2 phosphate buffer containing 1% polyvinylpyrrolidone, then tanned with 2% 5-sulphosalicylic acid in pH 7.2 phosphate buffer containing 1% polyvinylpyrrolidone, and collected by centrifugation.

FS CPI EPI

FA AB

MC CPI: A12-V03B; A12-V03C; B04-B04D; B12-K04

=> d his

(FILE 'HOME' ENTERED AT 12:39:18 ON 03 MAR 2006)

FILE 'HCAPLUS' ENTERED AT 12:39:28 ON 03 MAR 2006

L1 1 US2005287513/PN OR US2003-696023#/AP, PRN  
 E DAVIS A/AU

L2 224 E3, E35-36

E DAVIS ASHLEY/AU

L3 22 E3, E6-7

E MIDDLETON K/AU

L4 9 E3, E7

E MIDDLETON KIM/AU

L5 20 E3-5

37 CYTOSKELETON/CS, PA

FILE 'REGISTRY' ENTERED AT 12:41:50 ON 03 MAR 2006

FILE 'HCAPLUS' ENTERED AT 12:41:54 ON 03 MAR 2006

L7 TRA L1 1- RN : 4 TERMS

FILE 'REGISTRY' ENTERED AT 12:41:54 ON 03 MAR 2006

L8 4 SEA L7

L9 13322 CH4O

L10 QUE (PMS OR MAN OR IDS)/CI OR UNSPECIFIED OR COMPD OR COMPOUND

L11 1083 L9 NOT L10  
 L12 748 L11 NOT M/ELS  
 L13 88374 CSH8O2  
 L14 950 L13 NOT L10

FILE 'HCAPLUS' ENTERED AT 12:47:27 ON 03 MAR 2006

L15 133866 L12  
 L16 73262 L14  
 L17 209767 METHANOL OR BIELESKI (1A) SOLUT? OR CARBINOL OR METHYL (1W) (ALCO  
 L18 32909 GLUTARALDEHYDE OR PENTANEDIAL OR PENTANEDIONE OR ALDESAN OR AQU  
 L19 125 GLUTOHYDE OR HOSPEX OR KCIDE OR NSC13392 OR NSC () (13392 OR 13  
     E TISSUE CULTURE/CT  
     E E3+ALL  
     E E2+ALL  
 L20 37091 E1+OLD  
     E CELL CULTURE/CT  
     E E3+ALL  
     E E3+ALL  
 L21 24043 E3+OLD, NT  
     E PLANT CELL/CT  
     E E3+ALL  
 L22 57837 E2+OLD, NT  
     E PLANT CELL/CT  
 L23 8189 E3-9  
     E PLANT TISSUE/CT  
 L24 19081 E3-37  
     E E3+ALL  
 L25 24711 E1+OLD, NT  
     E PLANT TISSUE CULTURE/CT  
 L26 9490 E4-13  
 L27 830 L15-19 AND L20-26  
     E PRESERVATIVE/CT  
 L28 6453 E4-5  
     E E4+ALL  
 L29 109013 E2+NT  
 L30 17 L27 AND L28-29  
 L31 1 L30 AND L1-6  
 L32 16 L30 NOT L31  
     E ANIMAL CELL LINE/CT  
     E E3+ALL  
 L33 96724 E3+NT  
 L34 468 L33 AND L15-19  
 L35 9 L34 AND L28-29  
 L36 1 L35 AND L1-6  
 L37 23 L32, L35  
 L38 22 L37 NOT L31, L36  
     E CARBOHYDRATE/CT  
     E E3+ALL  
     E E2  
 L39 108216 E3-111  
     E E3+OLD, NT1  
 L40 282679 E1+OLD, NT1  
     E E63+ALL  
 L41 536250 E3+OLD, NT  
     E GLYCOCONJUGATES/CT  
 L42 1583 E3-4  
     E E3+ALL  
 L43 2818 E3+OLD, NT  
     E OLIGOSACCHARIDES/CT  
     E E3+AL  
     E E3+ALL  
 L44 186074 E3+OLD, NT  
     E OLIGOSACCHARIDES/CT  
 L45 20097 E4-75  
     E SIALIC ACIDS/CT  
 L46 9292 E3-11

E E3+ALL  
 L47 10862 E3+OLD, NT  
 E GLYCOSIDES/CT  
 L48 39965 E3-126  
 E E3+ALL  
 L49 581533 E3+OLD, NT  
 E CARBOHYDRATES+OLD, NT1/CT  
 E E70+ALL  
 L50 17860 E3+OLD, NT  
 E CYCLITOLS/CT  
 L51 1350 E3-5  
 E E3+ALL  
 L52 53043 E6+NT  
 E CYCLODEXTRIN/CT  
 L53 6081 E3-11  
 E E3+ALL  
 L54 8254 E3-4  
 E BUFFERS/CT  
 E E3+ALL  
 L55 12459 E1+OLD  
 L56 253 L27 AND L29-54  
 L57 9 L56 AND L55  
 L58 1 L57 AND L1-6  
 L59 1 L31, L36, L58  
 L60 1 L59 AND L1-6  
 L61 1 L59 AND L15-58  
 L62 1 L60-61  
 E PRESERVATION/CT  
 E PRESERVATION/CT  
 L63 3275 E3-7  
 E E3+ALL  
 L64 16313 E1+NT  
 L65 17 L64 AND L27  
 L66 1 L65 AND L1-6  
 L67 16 L65 NOT L66  
 SEL AN 8  
 L68 1 E1-2 AND L67  
 L69 2 L62, L66, L68

FILE 'BIOSIS' ENTERED AT 14:32:06 ON 03 MAR 2006

L70 52468 L15-19  
 L71 28 CYTOSKELETON/CS  
 E DAVIS A/AU  
 L72 405 E3, E29-32  
 E DAVIS ASHLEY/AU  
 L73 31 E3-6  
 E MIDDLETON K/AU  
 L74 81 E3-8  
 E MIDDLETON KIM/AU  
 L75 14 E3-4  
 L76 0 L70 AND L71-75

FILE 'WPIX' ENTERED AT 14:34:05 ON 03 MAR 2006

L77 417223 (C07C031-04 OR A61K031-045)/IPC OR H401/M0, M1, M2, M3, M4, M5, M6  
 E C07C031/IC, ICM, ICS  
 L78 1257 E35-37  
 E C07C031/ICA, ICI  
 L79 26 E7  
 E A61K031-045/IC, ICM, ICS  
 L80 2034 E3-7  
 E A61K031-045/ICA, ICI  
 L81 72 E3-4  
 E A61K031:045/ICA, ICI  
 L82 91 E4  
 E METHANOL/CN  
 L83 4 E3-7, E17-18

E GLUTARALDEHYDE/CN  
 L84 1 E3  
 L85 5 L83-84  
     SEL DCSE L85  
     EDIT /DCSE /DCRE  
 L86 4975 E1-5  
     SEL SDCN L85  
     EDIT /SDCN /DCN  
 L87 7020 E6-10  
 L88 22733 (0270 OR 0927)/DRN  
 L89 593 A61K031-11/IPC  
     E A61K031-11/IC, ICM, ICS  
 L90 575 E3-7  
     E A61K031-11/ICA, ICI  
 L91 20 E3-4  
     E A61K031:11/ICA, ICI  
 L92 5 E4  
 L93 86392 L17-19  
     E DAVIS A/AU  
 L103 97 E3, E24  
     E MIDDLETON K/AU  
 L104 10 E3, E6  
 L105 2 CYTOSKELET?/PA, CS  
 L106 0 CYTO/PACO  
 L107 590 CYTO-N/PACO  
 L108 1 L99 AND L103-107  
 L109 69 L99 AND L93  
 L110 27 L109 NOT (PY>2003 OR AY>2003 OR PRY>2003)  
     SEL AN 1 9 12 26 L110  
 L111 4 E1-4 AND L110  
 L112 39400 L77-82, L86-93 AND L100  
 L113 6762 L112 AND L97-98  
 L114 217 L94 AND L113  
 L115 217 L101, L114  
 L116 20 L115 AND L93  
     SEL AN 2 4 5 10 L116  
 L117 4 E5-8 AND L116  
 L118 13 L100 AND L97-98 AND L95 AND L94  
 L119 23038 L100 AND L97-98  
 L120 6762 L119 AND L77-82, L86-93  
 L121 103 L120 AND FIXING  
 L122 2 L121 AND L94  
 L123 9 L108, L111, L117  
 L124 9 L123 AND L77-82, L86-122

FILE 'WPIX' ENTERED AT 16:31:16 ON 03 MAR 2006

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